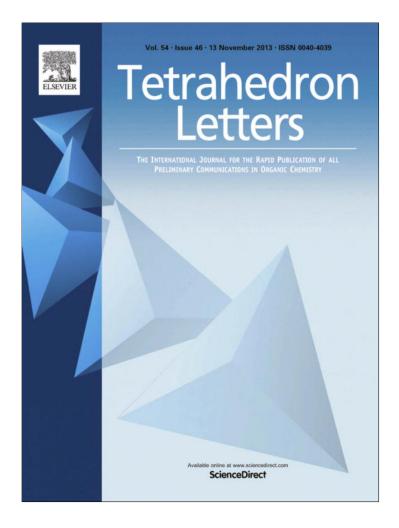
Provided for non-commercial research and education use. Not for reproduction, distribution or commercial use.



This article appeared in a journal published by Elsevier. The attached copy is furnished to the author for internal non-commercial research and education use, including for instruction at the authors institution and sharing with colleagues.

Other uses, including reproduction and distribution, or selling or licensing copies, or posting to personal, institutional or third party websites are prohibited.

In most cases authors are permitted to post their version of the article (e.g. in Word or Tex form) to their personal website or institutional repository. Authors requiring further information regarding Elsevier's archiving and manuscript policies are encouraged to visit:

http://www.elsevier.com/authorsrights

Tetrahedron Letters 54 (2013) 6181-6184

Contents lists available at ScienceDirect

Tetrahedron Letters

journal homepage: www.elsevier.com/locate/tetlet

Regulation of dynamic structure of cyclophanes by their complexation with the porphyrin



etrahedro

Akihiko Tsuge*, Masahiro Yokoo, Hiroyoshi Kawasaki, Daisaku Kaneko, Tetsuji Moriguchi, Koji Araki

Department of Applied Chemistry, Kyushu Institute of Technology, Tobata-ku, Kitakyushu 804-8550, Japan

ARTICLE INFO

Article history: Received 2 July 2013 Revised 20 August 2013 Accepted 26 August 2013 Available online 5 September 2013

Keywords: Cyclophane Porphyrin Conformation Complex

ABSTRACT

Dithia[3.3]metacyclophanes which consist of the pyridine unit connecting to the different positions of the parent cyclophane skeleton have been prepared. Conformational change has been observed for the cyclophane having a 4-substituted pyridine unit by binding to the porphyrin. In contrast the porphyrin binding has no influence on conformational behavior of the cyclophane having a 3-substituted pyridine unit. © 2013 Elsevier Ltd. All rights reserved.

Cyclophanes known as bridged aromatic compounds have been intensively investigated from various points of view because of their unique characteristics for the last few decades.¹ One of the most characteristic features of cyclophanes is their dynamic structure based on the conformational flexibility.² Thus, the conformational properties have been well studied in various cyclophane systems. Meanwhile we have been interested in the small-sized dithia[3.3]metacyclophane system because of their unique π systems owing to strong transannular π -electronic interactions between aromatic components in close proximity.³ These dithia[3.3]metacyclophanes exhibit conformational features based on their ring inversion between the syn and anti conformations.^{3i,4} Control of this conformational behavior is one of the attracting topics because some substantial characteristics of cyclophanes highly depend on their dynamic structures. On the other hand porphyrins are known to bind the pyridine derivatives.⁵

We have already reported the syntheses and properties of pyridinophanes that are the cyclophanes consisting of the pyridine component.⁶ In this respect we describe here the conformational regulation of pyridinophanes by complexation with the porphyrins.

Starting with 5-hydroxyisophthalic acid, esterification and reaction with chloromethyl pyridines, followed by reduction, and chloromethylation gave the chloromethyl compounds 1a-c. The sulfanylmethyl compounds 2a-c were obtained from 1a-c (see Fig. 1). The coupling reactions of 1a-c and 2a-c were carried out using Cs₂CO₃ as a base under a highly diluted condition to give

* Corresponding author. *E-mail address:* tsuge@che.kyutech.ac.jp (A. Tsuge).

0040-4039/\$ - see front matter @ 2013 Elsevier Ltd. All rights reserved. http://dx.doi.org/10.1016/j.tetlet.2013.08.116 the corresponding pyridinophanes 3a-c and 3e in the yields of 43-57% (see Fig. 2).

The ¹H NMR spectral titration has been carried out for **3a–c** with the zinc porphyrin **4a**. Figure 3 shows the profile of the ¹H NMR spectrum of **3a** on addition of **4a**.

The addition of **4a** induces the upfield shifts for the protons (H_A and H_B) neighboring the nitrogen atom, indicating that the binding of porphyryin against pyridinophane should occur. In over 1:3 ratio no remarkable shifts were observed. In accordance with this porphyrin binding the upfield shift of the inner proton (H_C) is observed as shown in Figure 3. It has been well known that an extensive upfield shift of the proton at the inner position in analogous [2.2]metacyclophanes is recognized due to the strong shielding effect of the opposite aromatic ring in the *anti* conformation.⁷ It has also been reported that there exists a rapid inversion between a *syn* conformation and an *anti* conformation in the dithia[3.3]metacyclophane system at room temperature.⁸ The addition of **4a** to the

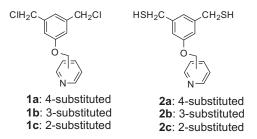


Figure 1. Chemical structures of 1 and 2.

Author's personal copy

A. Tsuge et al./Tetrahedron Letters 54 (2013) 6181-6184

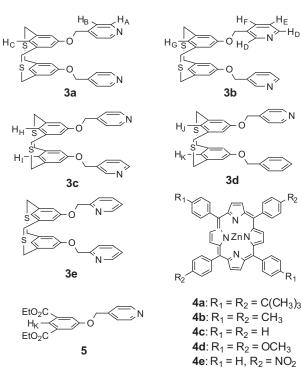


Figure 2. Chemical structures of 3, 4, and 5.

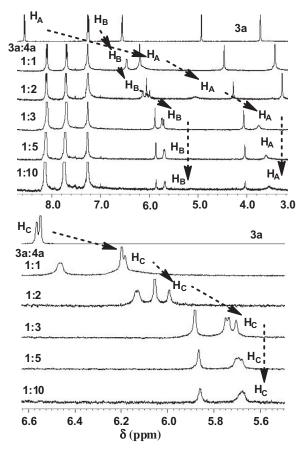


Figure 3. Partial NMR spectra of 3a on addition of 4a.

referential compound **5** under the same condition has little influence on the chemical shift of the inner proton (H_K) in **5**. Taking these results into account the upfield shift of the inner proton

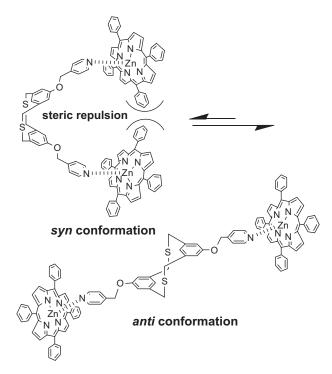


Figure 4. Preference for the *anti* conformation in the cyclophane–porphyrin system.

 (H_c) seen in Figure 3 strongly suggests that the contribution of the *anti* conformation increases on the addition of porphyrin. This preference for the *anti* conformation can be considered as consequences of hindrance for formation of the *syn* conformation by means of the bulky aromatic cyclophane component binding the porphyrin ring as illustrated in Figure 4.

A similar ¹H NMR spectral titration was done for the pyridinophane **3b** and **4a** as shown in Figure 5.

The binding of porhyrin to **3b** can also be confirmed by the upfield shifts for the protons (H_D , H_E , and H_F) on addition of **4a**, however, the chemical shift of the inner proton (H_G) shows almost no change in contrast to **3a**. No chemical shift of the inner proton in the pyridinophane **3b** indicates that the binding of porphyrin to **3b** might have little effect on its conformational inversion, meaning that the ring inversion takes place without bumping between two bulky cyclophane components attached by the porphyrin rings. This should be due to the binding of porphyrin on the position of 3-substituted nitrogen in pyridinophane.

In order to confirm this assumption we have also examined the ¹H NMR spectral titration employing the pyridinophane **3c** and **3d** as shown in Figure 6. The inner protons (H_H and H_I) in **3c** exhibit about 0.45 ppm upfield shift on addition of the porphyrin **4a**. Under similar condition the upfield shifts for the inner protons (H_J and H_K) in **3d** are 0.33 ppm and 0.27 ppm, respectively. These values are smaller than the corresponding shift (0.88 ppm) recognized for **3a**, indicating that the *anti* conformation makes less contribution to the process of inversion between the *syn* and *anti* conformations compared with **3a**.

These results strongly suggest the bulky porphyrin ring on the position of 3-substituted nitrogen has a slight effect on the ring inversion in pyridinophanes. No apparent binding of **4a** against the pyridinophane **3e** was observed probably due to the steric hindrance.

Temperature should be considered for binding of porphyrin to pyridine derivatives. Thus, we have examined the conformational behavior of the porphyrin-bound pyridinophane **3a** depending on the temperature as shown in Figure 7.

6182

A. Tsuge et al./Tetrahedron Letters 54 (2013) 6181-6184

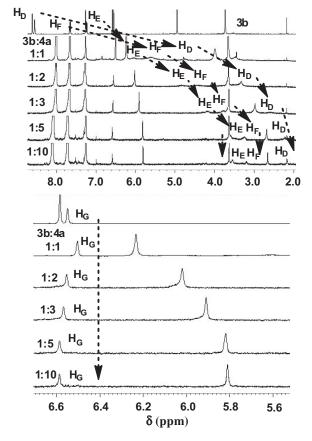


Figure 5. Partial NMR spectra of 3b on addition of 4a.

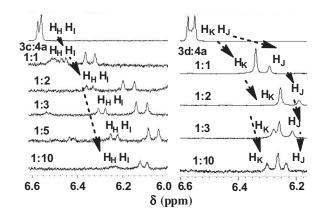


Figure 6. Partial NMR spectra of 3c and 3d on addition of 4a.

There was observed an upfield shift of the inner proton (H_C) in accordance with the shifts of the protons (H_A and H_B) as the temperature was going down to -30 °C. With more cooling down the corresponding peaks disappeared owing to an extensive broadening. The upfield shifts of the protons (H_A and H_B) mean there is an enhancement in binding of porphyrin toward pyridinophane **3a** when the temperature goes down. A preference for the *anti* conformation indicated by the chemical shift of the inner proton (H_C) can be ascribed to an enhanced binding of porphyrin. In contrast raising the temperature the NMR profile is apparently becoming close to that of the pyridinophane **3a** itself, meaning that the process of the ring inversion in the porphyrin-bound pyridinophanes is approaching the original conformational change of **3a**. Increasing

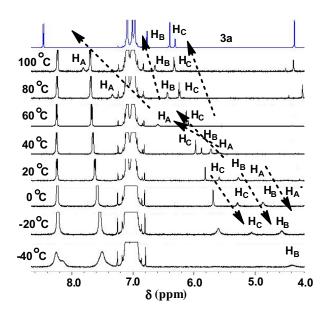


Figure 7. Partial VT spectra of 3a and 4a.

the temperature is considered to promote a dissociation of porphyrin from the porphyrin-bound pyridinophanes. From this point of view it can be confirmed that movement to *anti* conformation in the conformational exchange of pyridinophanes is triggered by complexation of porphyrin toward pyridinophanes.

The observations obtained here imply that the binding of porphyrin regulates the ring inversion of pyridinophanes. This binding should be dependent on the structure of porphyrins. Thus, some porphyrin derivatives (4b-e) have been prepared and the association constants in their binding against pyridinophanes have been calculated by the NMR titrations as summarized in Table 1.

The chemical shift changes of the inner protons upon addition of porphyrin are also inserted in Table 1. The association constants obtained for 3a with 4a show the same order as to those for 3a with 4b, 4c, and 4d. In these cases the chemical shift changes of the inner proton in 3a on addition of 4a, 4b, 4c, and 4d are from 0.88 to 1.23 ppm. A large association constant is observed for 3a and **4e**, which can be ascribed to the nitro group on the porhyrin working as an electron-withdrawing group. This large association constant seems to be consistent with the largest upfield shift of the inner proton. This fact also supports the assumption that the binding of porphyrin should hamper the syn conformation, resulting in preference for the anti conformation. The association constants of the same order of $10^7 \, \text{M}^{-2}$ for **3b** with **4a-e** were confirmed as for **3a**. These results well agree with strong binding of porphyrin against **3b** anticipated by the NMR titration. In this case the largest association constant was likewise obtained for nitro porphyrin 4e. Contrary to 3a no obvious chemical shifts of the

Table 1
Association constants ^a and chemical shift changes ^b for 3a and 3b with 4a-4e

		Association constant (K/M ⁻²)			
	3a		3b		
4a	$\textbf{4.0}\times\textbf{10}^{7}$	(0.88 ppm)	$\textbf{4.0}\times \textbf{10}^{7}$	(0 ppm)	
4b	$5.6 imes 10^7$	(0.89 ppm)	$1.4 imes 10^7$	(0.02 ppm)	
4c	$1.3 imes 10^7$	(0.94 ppm)	$1.3 imes 10^7$	(0.02 ppm)	
4d	$3.8 imes 10^7$	(1.23 ppm)	$1.2 imes 10^7$	(0.02 ppm)	
4e	$1.2 imes 10^7$	(1.42 ppm)	$\textbf{9.4}\times 10^7$	(0.08 ppm)	

^a In chloroform, T = 295 K.

^b Values are in parenthesis.

inner protons in **3b** on addition of porphyrins were recognized except for a very slight shift observed for addition of **4e**. These results strongly indicate that the binding of porphyrin at 3-substituted nitrogen in pyridinophane has nothing to do with restriction of its ring inversion.

It should be noted that conformational regulation from a *syn* conformation to an *anti* conformation in the [3.3]metacyclophane system consisting of pyridine units has been achieved by external stimuli such as addition of porhyrin. This regulation is strongly dependent on the position of substituted pyridine unit to which porphyrin binds.

Supplementary data

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.tetlet.2013. 08.116.

References and notes

- For reviews, see: (a) Smith, B. H. Bridged Aromatic Compounds; Academic Press: New York, 1964; (b)Cyclophanes; Keehn, P. M., Rosenfeld, S. M., Eds.; Academic Press: New York, 1983. Vols. 1 and 2; (c) Diederich, F. Cyclophanes; Royal Society of Chemistry: London, 1991; (d) Vögtle, F. Cyclophane Chemistry; Wiley: New York, 1993; (e) Kane, V. V.; de Wolf, W. H.; Bickelhaupt, F. Tetrahedron 1994, 50, 4575; (f) Bodwell, G. J. Angew. Chem., Int. Ed. Engl. 1996, 35, 2085; (g) de Maijere, A.; König, B. Synlett 1997, 1221; (h) Bodwell, G. J. In Organic Synthesis Highlights; Schmalz, H. G., Ed.; Wiley-VCH: New York, 2000; Vol. IV, (i) Hopf, H. Classics in Hydrocarbon Chemistry; Wiley-VCH: Weinheim, 2000; (j)Modern Cyclophane Chemistry; Gleiter, R., Hopf, H., Eds.; Wiley-VCH: Weinheim, 2004.
- (a) Mitchell, R. H. In *Cyclophanes*; Keehn, P. M., Rosenfeld, S. M., Eds.; Academic Press: New York, 1983; Vol. I, (b) Moriguchi, T.; Sakata, K.; Tsuge, A. *Chem. Lett.* 1999, 167; (c) Nakamura, Y.; Fujii, T.; Sugita, S.; Nishimura, J. *Chem. Lett.* 1999, 1039; (d) Shull, B. K.; Spielvogel, D. E.; Gopalaswamy, R.; Sankar, S.; Boyle, P. D.;

Head, G.; Devito, K. J. Chem. Soc., Perkin Trans. 2 **2000**, 557; (e) Laali, K. K.; Borodkin, G. I.; Okazaki, T.; Hayashida, Y.; Nakamura, Y.; Nishimura, J. J. Chem. Soc., Perkin Trans. 2 **2000**, 2347; (f) Moriguchi, T.; Sakata, K.; Tsuge, A. J. Chem. Soc., Perkin Trans. 2 **2001**, 934; (g) Isaji, H.; Yasutake, M.; Takemura, H.; Sako, K.; Tatemitsu, H.; Inazu, T.; Shinmyozu, T. *Eur. J. Org. Chem.* **2001**, 2487; (h) Mitchell, R. H. J. Am. Chem. Soc. **2002**, 124, 2352; (i) Bodwell, G. J.; Li, J. Org. Lett. **2002**, 4, 127; (j) Yamato, T.; Tsuchihashi, K.; Nakamura, N.; Hirahara, M.; Tanaka, K. Can. J. Chem. **2002**, 80, 510.

- (a) Tsuge, A.; Araki, T.; Noguchi, Y.; Yasutake, M.; Moriguchi, T.; Sakata, K. Chem. Lett. 1998, 603; (b) Tsuge, A.; Nishimoto, T.; Uchida, T.; Yasutake, M.; Moriguchi, T.; Sakata, K. J. Org. Chem. 1999, 64, 7246; (c) Tsuge, A.; Nada, R.; Moriguchi, T.; Sakata, K. J. Org. Chem. 2001, 66, 9023; (d) Tsuge, A.; Tanba, Y.; Moriguchi, T.; Sakata, K. Chem. Lett. 2002, 384; (e) Tsuge, A.; Nishimoto, H.; Moriguchi, T.; Sakata, K. Chem. Lett. 2003, 32, 776; (f) Tsuge, A.; Iwasaki, W.; Moriguchi, T.; Sakata, K. Chem. Lett. 2004, 33, 756; (g) Tsuge, A.; Ikeura, Y.; Moriguchi, T.; Sakata, K. Org. Biomol. Chem. 2005, 3, 3590; (h) Tsuge, A.; Ikeura, Y.; Moriguchi, T.; Yamaji, M. Chem. Lett. 2008, 37, 870; (j) Tsuge, A.; Hara, T.; Moriguchi, T. Tetrahedron Lett. 2009, 50, 4509.
- (a) Moriguchi, T.; Sakata, K.; Tsuge, A. Chem. Lett. **1999**, 167; (b) Moriguchi, T.; Sakata, K.; Tsuge, A. J. Chem. Res. Synop. **1999**, 604.
 (a) Iengo, E.; Zangrando, E.; Alessio, E. Eur. J. Inorg. Chem. **2003**, 2371; (b) Slagt, V.
- (a) lengo, E.; Zangrando, E.; Alessio, E. *Eur. J. Inorg. Chem.* **2003**, 2371; (b) Slagt, V. F.; Leeuwen, P. W. N. M.; Reek, J. N. H. *Dalton Trans.* **2007**, 2302; (c) Wu, J.; Fang, F.; Lu, W. H.; Hou, J. L.; Li, C.; Wu, Z. Q.; Jiang, X. Y.; Li, Z. T.; Yu, Y. H. *J. Org. Chem.* **2007**, 72, 2897; (d) Aratani, N.; Kim, D.; Osuka, A. *Acc. Chem. Res.* **2009**, 42, 1922.
 Tsuge, A.; Matsubara, A.; Moriguchi, T.; Sei, Y.; Yamaguchi, K. *Tetrahedron Lett.*
- Tsuge, A.; Matsubara, A.; Moriguchi, T.; Sei, Y.; Yamaguchi, K. *Tetrahedron Lett.* 2006, 47, 6607.
 (a) Mitchell, R. H.; Weerawarna, K. S.; Bushnell, G. W. *Tetrahedron Lett.* 1984, 25,
- 907; (b) Mitchell, R. H.; Vindo, T. K.; Bushnell, G. W. J. Am. Chem. Soc. **1985**, 107, 3340; (c) Nishimura, J.; Horikoshi, Y.; Wada, Y.; Takahashi, H.; Sato, M. J. Am. Chem. Soc. **1991**, 113, 3485; (d) Caramori, G. F.; Galembeck, S. E.; Laali, K. K. J. Org. Chem. **2005**, 70, 3242.
- (a) Fukazawa, Y.; Takeda, Y.; Usui, S.; Kodama, M. J. Am. Chem. Soc. 1988, 110, 7842; (b) Albert, B.; Jansen, M.; Guther, R.; Vogtle, F. Acta Crystallogr. C 1993, 49, 2002; (c) Barclay, T.; Cordes, W.; Yang, F. M.; Lin, S. T. J. Chem. Crystallogr. 1997, 27, 359; (d) Bodwell, G. J.; Bridson, J. N.; Houghton, T. J.; Yarlagadda, B. Tetrahedron Lett. 1997, 38, 7475; (e) Yamato, T.; Tsuchihashi, K.; Nakamura, N.; Hirahara, M.; Tanaka, K. Can. J. Chem. 2002, 80, 510.

6184